**Zebrafish Injections**

1. Check labelling of cells
	1. Brightfield on scope 🡪 adjust light on the bottom of the stage and adjust stage
	2. Parafilm with nothing on 🡪 pipette on 5µL of cells (resusupend) 🡪 focus on drop
	3. Assign labels to makers for both 🡪 sample can be reused after!
2. Make tricane
	1. Dilute stock 1:1000 in E3
	2. Make 30mLish
3. **Put on needle**
	1. Plunger needs to be in the upright position
	2. Use forceps to cut needle as needed
	3. Flow mineral oil through with syringe (use dot to check the diameter of the needle 🡪 important for consistent output)
	4. Put around plunger and tighten black twisty 🡪 give needle a little tug to check
	5. Make sure there are no bubbles made sure plunger is all the way up
	6. Empty until there is little oil left
4. Get agarose gel ready and 2 normal plates (one with dilute tricane and one with E3 for recovery)
5. 3 dpf zebrafish (10nL injection set 🡪 depends on needle diameter
6. Parafilm on microinjector stage and pipette on 5µL of cells
7. Suck up some cells with “fill” button and empty a little on a kim wipe so clear out any air
8. Anesthetise fish in tricane for about 1 min until they stop swimming
	1. Need to have a little tricane on the agarose
9. Inject at about a 45° angle 🡪 steeper the better!
10. INJECT!
11. Check injections under fluorescent scope
12. E3 for recovery!